

# Package ‘scPOEM’

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**Type** Package

**Title** Single-Cell Meta-Path Based Omic Embedding

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**Description** Provide a workflow to jointly embed chromatin accessibility peaks and expressed genes into a shared low-dimensional space using paired single-cell ATAC-seq (scATAC-seq) and single-cell RNA-seq (scRNA-seq) data. It integrates regulatory relationships among peak-peak interactions (via 'Cicero'), peak-gene interactions (via Lasso, random forest, and XGBoost), and gene-gene interactions (via principal component regression). With the input of paired scATAC-seq and scRNA-seq data matrices, it assigns a low-dimensional feature vector to each gene and peak. Additionally, it supports the reconstruction of gene-gene network with low-dimensional projections (via epsilon-NN) and then the comparison of the networks of two conditions through manifold alignment implemented in 'scTenifoldNet'.

**License** GPL (>= 2)

**Encoding** UTF-8

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align\_embedding      *Gene Network Reconstruction and Alignment*

### Description

Reconstruct gene networks via epsilon-NN and compare conditions using manifold alignment implemented in scTenifoldNet.

### Usage

```
align_embedding(
  gene_data1,
  gene_node1,
  E1,
  gene_data2,
  gene_node2,
  E2,
  dirpath = tempdir(),
  save_file = TRUE,
  d = 100
)
```

### Arguments

|            |   |
|------------|---|
| gene_data1 | The information for genes in state1, must have a col names "gene_name". |
| gene_node1 | Gene ids that are associated with other peaks or genes in state1.       |
| E1         | Embedding representations of peaks and genes in state1.                 |
| gene_data2 | The information for genes in state2, must have a col names "gene_name". |
| gene_node2 | Gene ids that are associated with other peaks or genes in state2.       |
| E2         | Embedding representations of peaks and genes in state2.                 |

|           |  |
|-----------|--|
| dirpath   | The folder path to read or write file          |
| save_file | Logical, whether to save the output to a file. |
| d         | The dimension of latent space.                 |

**Value**

A list containing the following elements:

- E\_g2 Low-dimensional embedding representations of genes under the two conditions.
- common\_genes Genes shared between both conditions and used in the analysis.
- diffRegulation A list of differential regulatory information for each gene.

**Examples**

```

library(scPOEM)
library(monocle)
dirpath <- "./example_data"
# Download compare mode example data
data(example_data_compare)
data_S1 <- example_data_compare$S1
data_S2 <- example_data_compare$S2
gg_net1 <- GGN(data_S1$Y, file.path(dirpath, "compare/S1"), save_file=FALSE)
pp_net1 <- PPN(data_S1$X, data_S1$peak_data, data_S1$cell_data,
                 data_S1$genome, file.path(dirpath, "compare/S1"), save_file=FALSE)

net_Lasso1 <- PGN_Lasso(data_S1$X, data_S1$Y,
                         data_S1$gene_data, data_S1$neibor_peak,
                         file.path(dirpath, "compare/S1"), save_file=FALSE)
net_RF1 <- PGN_RF(data_S1$X, data_S1$Y, data_S1$gene_data,
                     data_S1$neibor_peak, file.path(dirpath, "compare/S1"), save_file=FALSE)
net_XGB1 <- PGN_XGBoost(data_S1$X, data_S1$Y,
                         data_S1$gene_data, data_S1$neibor_peak,
                         file.path(dirpath, "compare/S1"), save_file=FALSE)
pg_net_list1 <- list(net_Lasso1, net_RF1, net_XGB1)
E_result_S1 <- pg_embedding(gg_net1, pp_net1, pg_net_list1,
                             file.path(dirpath, "compare/S1"), save_file=FALSE)

gg_net2 <- GGN(data_S2$Y, file.path(dirpath, "compare/S2"), save_file=FALSE)
pp_net2 <- PPN(data_S2$X, data_S2$peak_data,
                 data_S2$cell_data, data_S2$genome,
                 file.path(dirpath, "compare/S2"), save_file=FALSE)
net_Lasso2 <- PGN_Lasso(data_S2$X, data_S2$Y,
                         data_S2$gene_data, data_S2$neibor_peak,
                         file.path(dirpath, "compare/S2"), save_file=FALSE)
net_RF2 <- PGN_RF(data_S2$X, data_S2$Y, data_S2$gene_data,
                     data_S2$neibor_peak, file.path(dirpath, "compare/S2"), save_file=FALSE)
net_XGB2 <- PGN_XGBoost(data_S2$X, data_S2$Y,
                         data_S2$gene_data, data_S2$neibor_peak,
                         file.path(dirpath, "compare/S2"), save_file=FALSE)
pg_net_list2 <- list(net_Lasso2, net_RF2, net_XGB2)
E_result_S2 <- pg_embedding(gg_net2, pp_net2, pg_net_list2,
```

```

file.path(dirpath, "compare/S2"), save_file=FALSE)

compare_result <- align_embedding(data_S1$gene_data,
                                   E_result_S1$gene_node,
                                   E_result_S1$E,
                                   data_S2$gene_data,
                                   E_result_S2$gene_node,
                                   E_result_S2$E,
                                   file.path(dirpath, "compare/compare"),
                                   save_file=FALSE)

```

**eNN***Network Reconstruction via epsilon-NN***Description**

Reconstruction of gene-gene network via low-dimentional projections (via epsilon-NN).

**Usage**

```
eNN(E_g)
```

**Arguments**

|     |                                     |
|-----|-------------------------------------|
| E_g | Embedding representations of genes. |
|-----|-------------------------------------|

**Value**

The epsilon-NN network.

**example\_data\_compare    Example Input Data for Compare Mode Analysis****Description**

A list containing example single-cell multi-omics data used in "compare" mode of the scPOEM package.

**Usage**

```
data(example_data_compare)
```

## Format

A named list of length 2. Each element is itself a named list with the following components:

- X The scATAC-seq data, sparse matrix.
- Y The scRNA-seq data, sparse matrix.
- peak\_data A data.frame containing peak information.
- gene\_data A data.frame containing gene information (must contain column "gene\_name").
- cell\_data A data.frame containing cell metadata.
- neibor\_peak The peak IDs within a certain range of each gene, must have cols c("gene\_name", "start\_use", "end\_use"). The id numbers in "start\_use" and "end\_use" are start from 0.
- genome The genome length for the species.

## Examples

```
data(example_data_compare)
```

---

example\_data\_single    *Example Input Data for Single Mode Analysis*

---

## Description

A list containing example single-cell multi-omics data used in "single" mode of the scPOEM package.

## Usage

```
data(example_data_single)
```

## Format

A named list with 7 elements:

- X The scATAC-seq data, sparse matrix.
- Y The scRNA-seq data, sparse matrix.
- peak\_data A data.frame containing peak information.
- gene\_data A data.frame containing gene information (must contain column "gene\_name").
- cell\_data A data.frame containing cell metadata.
- neibor\_peak The peak IDs within a certain range of each gene, must have cols c("gene\_name", "start\_use", "end\_use"). The id numbers in "start\_use" and "end\_use" are start from 0.
- genome The genome length for the species.

## Examples

```
data(example_data_single)
```

GGN

*Construct Gene-Gene Network***Description**

Construct the gene-gene network via principle component regression.

**Usage**

```
GGN(
  Y,
  dirpath = tempdir(),
  count_device = 1,
  nComp = 5,
  rebuild_GGN = TRUE,
  save_file = TRUE,
  python_env = "scPOEM_env"
)
```

**Arguments**

|              |  |
|--------------|--|
| Y            | The scRNA-seq data, sparse matrix.   |
| dirpath      | The folder path to read or write file.   |
| count_device | The number of cpus used to train the Lasso model.  |
| nComp        | The number of PCs used for regression  |
| rebuild_GGN  | Logical. Whether to rebuild the gene-gene network (GGN) from scratch. If FALSE, the function will attempt to read from GGN.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode. |
| save_file    | Logical, whether to save the output to a file.   |
| python_env   | Name or path of the Python environment to be used.   |

**Value**

The GGN network.

**Examples**

```
library(scPOEM)
dirpath <- "./example_data"
# Download single mode example data
data(example_data_single)
# Construct GGN net.
gg_net <- GGN(example_data_single$Y,
               file.path(dirpath, "single"),
               save_file=FALSE)
```

---

|                  |                                    |
|------------------|------------------------------------|
| <b>PGN_Lasso</b> | <i>Peak-Gene Network via Lasso</i> |
|------------------|------------------------------------|

---

**Description**

Construct the peak-gene network via Lasso.

**Usage**

```
PGN_Lasso(
  X,
  Y,
  gene_data,
  neibor_peak,
  dirpath = tempdir(),
  count_device = 1,
  rebuild_PGN_Lasso = TRUE,
  save_file = TRUE
)
```

**Arguments**

|                   |  |
|-------------------|--|
| X                 | The scATAC-seq data, sparse matrix.  |
| Y                 | The scRNA-seq data, sparse matrix.   |
| gene_data         | The information for genes, must have a col names "gene_name".  |
| neibor_peak       | The peak IDs within a certain range of each gene, must have cols c("gene_name", "start_use", "end_use"). The id numbers in "start_use" and "end_use" are start from 0.   |
| dirpath           | The folder path to read or write file.   |
| count_device      | The number of cpus used to train the Lasso model.  |
| rebuild_PGN_Lasso | Logical. Whether to rebuild the peak-gene network via Lasso from scratch. If FALSE, the function will attempt to read from PGN_Lasso.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode. |
| save_file         | Logical, whether to save the output to a file.   |

**Value**

The PGN\_Lasso network.

**Examples**

```
library(scPOEM)
dirpath <- "./example_data"
# Download single mode example data
data(example_data_single)
```

```
# Construct PGN net via Lasso.
net_Lasso <- PGN_Lasso(example_data_single$X,
                        example_data_single$Y,
                        example_data_single$gene_data,
                        example_data_single$neibor_peak,
                        file.path(dirpath, "single"),
                        save_file=FALSE)
```

## PGN\_RF

*Peak-Gene Network via Random Forest***Description**

Construct the peak-gene network via random forest.

**Usage**

```
PGN_RF(
  X,
  Y,
  gene_data,
  neibor_peak,
  dirpath = tempdir(),
  count_device = 1,
  rebuild_PGN_RF = TRUE,
  save_file = TRUE,
  seed = NULL,
  python_env = "scPOEM_env"
)
```

**Arguments**

|                |   |
|----------------|---|
| X              | The scATAC-seq data, sparse matrix.   |
| Y              | The scRNA-seq data, sparse matrix.  |
| gene_data      | The information for genes, must have a col names "gene_name".   |
| neibor_peak    | The peak IDs within a certain range of each gene, must have cols c("gene_name", "start_use", "end_use"). The id numbers in "start_use" and "end_use" are start from 0.  |
| dirpath        | The folder path to read or write file.  |
| count_device   | The number of cpus used to train the Lasso model.   |
| rebuild_PGN_RF | Logical. Whether to rebuild the peak-gene network via random forest from scratch. If FALSE, the function will attempt to read from PGN_RF.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode. |

|            |   |
|------------|---|
| save_file  | Logical, whether to save the output to a file.                        |
| seed       | An integer specifying the random seed to ensure reproducible results. |
| python_env | Name or path of the Python environment to be used.                    |

**Value**

The PGN\_RF network.

**Examples**

```
library(scPOEM)
dirpath <- "./example_data"
# Download single mode example data
data(example_data_single)
# Construct PGN net via random forest (RF).
net_RF <- PGN_RF(example_data_single$X,
                   example_data_single$Y,
                   example_data_single$gene_data,
                   example_data_single$neibor_peak,
                   file.path(dirpath, "single"),
                   save_file=FALSE)
```

**Description**

Construct the peak-gene network via XGBoost.

**Usage**

```
PGN_XGBoost(
  X,
  Y,
  gene_data,
  neibor_peak,
  dirpath = tempdir(),
  count_device = 1,
  rebuild_PGN_XGB = TRUE,
  save_file = TRUE
)
```

### Arguments

|                 |   |
|-----------------|---|
| X               | The scATAC-seq data, sparse matrix.   |
| Y               | The scRNA-seq data, sparse matrix.  |
| gene_data       | The information for genes, must have a col names "gene_name".   |
| neibor_peak     | The peak IDs within a certain range of each gene, must have cols c("gene_name", "start_use", "end_use"). The id numbers in "start_use" and "end_use" are start from 0.  |
| dirpath         | The folder path to read or write file.  |
| count_device    | The number of cpus used to train the Lasso model.   |
| rebuild_PGN_XGB | Logical. Whether to rebuild the peak-gene network via XGBoost from scratch.<br>If FALSE, the function will attempt to read from PGN_XGB.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode. |
| save_file       | Logical, whether to save the output to a file.  |

### Value

The PGN\_XGBoost network.

### Examples

```
library(scPOEM)
dirpath <- "./example_data"
# Download single mode example data
data(example_data_single)
# Construct PGN net via XGBoost.
net_XGB <- PGN_XGBoost(example_data_single$X,
                        example_data_single$Y,
                        example_data_single$gene_data,
                        example_data_single$neibor_peak,
                        file.path(dirpath, "single"),
                        save_file=FALSE)
```

### Description

Learn the low-dimensional representations for peaks and genes with a meta-path based method.

**Usage**

```
pg_embedding(
  gg_net,
  pp_net,
  pg_net_list,
  dirpath = tempdir(),
  relearn_pg_embedding = TRUE,
  save_file = TRUE,
  d = 100,
  numwalks = 5,
  walklength = 3,
  epochs = 100,
  neg_sample = 5,
  batch_size = 32,
  weighted = TRUE,
  exclude_pos = FALSE,
  seed = NULL,
  python_env = "scPOEM_env"
)
```

**Arguments**

|                      |  |
|----------------------|--|
| gg_net               | The gene-gene network.   |
| pp_net               | The peak-peak network.   |
| pg_net_list          | A list of peak-gene networks, constructed via different methods.   |
| dirpath              | The folder path to read or write file.   |
| relearn_pg_embedding | Logical. Whether to relearn the low-dimensional representations for peaks and genes from scratch. If FALSE, the function will attempt to read from node_embeddings.mtx, node_used_peak.csv, node_used_gene.csv under dirpath/embedding in single mode or dirpath/state_name/embedding in compare mode. |
| save_file            | Logical, whether to save the output to a file.   |
| d                    | Dimension of the latent space. Default is 100.   |
| numwalks             | Number of random walks per node. Default is 5.   |
| walklength           | Length of walk depth. Default is 3.  |
| epochs               | Number of training epochs. Default is 100.   |
| neg_sample           | Number of negative samples per positive sample. Default is 5.  |
| batch_size           | Batch size for training. Default is 32.  |
| weighted             | Whether the sampling network is weighted. Default is TRUE.   |
| exclude_pos          | Whether to exclude positive samples from negative sampling. Default is FALSE.  |
| seed                 | An integer specifying the random seed to ensure reproducible results.  |
| python_env           | Name or path of the Python environment to be used.   |

**Value**

A list containing the following:

- E Low-dimensional representations of peaks and genes
- peak\_node Peak ids that are associated with other peaks or genes.
- gene\_node Gene ids that are associated with other peaks or genes.

**Examples**

```
library(scPOEM)
library(monocle)
dirpath <- "./example_data"
# Download single mode example data
data(example_data_single)
gg_net <- GGN(example_data_single$Y,
               file.path(dirpath, "single"),
               save_file=FALSE)
pp_net <- PPN(example_data_single$X, example_data_single$peak_data,
               example_data_single$cell_data, example_data_single$genome,
               file.path(dirpath, "single"), save_file=FALSE)
net_Lasso <- PGN_Lasso(example_data_single$X, example_data_single$Y,
                       example_data_single$gene_data, example_data_single$neibor_peak,
                       file.path(dirpath, "single"), save_file=FALSE)
net_RF <- PGN_RF(example_data_single$X, example_data_single$Y,
                  example_data_single$gene_data, example_data_single$neibor_peak,
                  file.path(dirpath, "single"), save_file=FALSE)
net_XGB <- PGN_XGBoost(example_data_single$X, example_data_single$Y,
                        example_data_single$gene_data, example_data_single$neibor_peak,
                        file.path(dirpath, "single"), save_file=FALSE)
E_result <- pg_embedding(gg_net, pp_net, list(net_Lasso, net_RF, net_XGB),
                           file.path(dirpath, "single"), save_file=FALSE)
```

**Description**

Construct peak-peak network.

**Usage**

```
PPN(
  X,
  peak_data,
  cell_data,
  genome,
```

```

    dirpath = tempdir(),
    rebuild_PPN = TRUE,
    save_file = TRUE,
    seed = NULL
)

```

## Arguments

|             |  |
|-------------|--|
| X           | The scATAC-seq data, sparse matrix.  |
| peak_data   | The information for peaks, must have a col names "peak_name".  |
| cell_data   | The information for cells, must have a col names "cell_name".  |
| genome      | The genome length for the species.   |
| dirpath     | The folder path to read or write file.   |
| rebuild_PPN | Logical. Whether to rebuild the peak-peak network (PPN) from scratch. If FALSE, the function will attempt to read from PPN.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode. |
| save_file   | Logical, whether to save the output to a file.   |
| seed        | An integer specifying the random seed to ensure reproducible results.  |

## Value

The PPN network.

## Examples

```

library(scPOEM)
library(monocle)
dirpath <- "./example_data"
# Download single mode example data
data(example_data_single)
# Construct PPN net.
pp_net <- PPN(example_data_single$X,
                example_data_single$peak_data,
                example_data_single$cell_data,
                example_data_single$genome,
                file.path(dirpath, "single"),
                save_file=FALSE)

```

## Description

This function takes paired single-cell ATAC-seq (scATAC-seq) and RNA-seq (scRNA-seq) data to embed peaks and genes into a shared low-dimensional space. It integrates regulatory relationships from peak-peak interactions (via Cicero), peak-gene interactions (via Lasso, random forest, and XGBoost), and gene-gene interactions (via principal component regression). Additionally, it supports gene-gene network reconstruction using epsilon-NN projections and compares networks across conditions through manifold alignment (scTenifoldNet).

## Usage

```
scPOEM(
  mode = c("single", "compare"),
  input_data,
  dirpath = tempdir(),
  count_device = 1,
  nComp = 5,
  seed = NULL,
  numwalks = 5,
  walklength = 3,
  epochs = 100,
  neg_sample = 5,
  batch_size = 32,
  weighted = TRUE,
  exclude_pos = FALSE,
  d = 100,
  rebuild_GGN = TRUE,
  rebuild_PPN = TRUE,
  rebuild_PGN_Lasso = TRUE,
  rebuild_PGN_RF = TRUE,
  rebuild_PGN_XGB = TRUE,
  relearn_pg_embedding = TRUE,
  save_file = TRUE,
  pg_method = c("Lasso", "RF", "XGBoost"),
  python_env = "scPOEM_env"
)
```

## Arguments

|                         |  |
|-------------------------|--|
| <code>mode</code>       | The mode indicating whether to analyze data from a single condition or to compare two conditions.  |
| <code>input_data</code> | <p>A list of input data.</p> <p>If <code>mode = "single"</code>, <code>input_data</code> must be a list containing the following <b>seven objects</b>:</p> <ul style="list-style-type: none"> <li>• <code>X</code>: The scATAC-seq data, sparse matrix.</li> <li>• <code>Y</code>: The scRNA-seq data, sparse matrix.</li> <li>• <code>peak_data</code>: A data.frame containing peak information.</li> <li>• <code>gene_data</code>: A data.frame containing gene information (must contain a column "gene_name").</li> </ul> |

- **cell\_data**: A data.frame containing cell metadata.
- **neibor\_peak**: The peak IDs within a certain range of each gene, must have cols c("gene\_name", "start\_use", "end\_use"). The id numbers in "start\_use" and "end\_use" are start from 0.
- **genome**: The genome length for the species.

If mode = "compare", input\_data must be a **named list of two elements**, with names corresponding to two state names (e.g., "S1" and "S2"). Each element must itself be a list containing the same seven components as described above for mode = "single".

|                                   |  |
|-----------------------------------|--|
| <code>dirpath</code>              | The folder path to read or write file.   |
| <code>count_device</code>         | The number of cpus used to train models.   |
| <code>nComp</code>                | The number of PCs used for regression in constructing GGN.   |
| <code>seed</code>                 | An integer specifying the random seed to ensure reproducible results.  |
| <code>numwalks</code>             | Number of random walks per node. Default is 5.   |
| <code>walklength</code>           | Length of walk depth. Default is 3.  |
| <code>epochs</code>               | Number of training epochs. Default is 100.   |
| <code>neg_sample</code>           | Number of negative samples per positive sample. Default is 5.  |
| <code>batch_size</code>           | Batch size for training. Default is 32.  |
| <code>weighted</code>             | Whether the sampling network is weighted. Default is TRUE.   |
| <code>exclude_pos</code>          | Whether to exclude positive samples from negative sampling. Default is FALSE.  |
| <code>d</code>                    | The dimension of latent space. Default is 100.   |
| <code>rebuild_GGN</code>          | Logical. Whether to rebuild the gene-gene network from scratch. If FALSE, the function will attempt to read from GGN.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode.   |
| <code>rebuild_PPN</code>          | Logical. Whether to rebuild the peak-peak network from scratch. If FALSE, the function will attempt to read from PPN.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode.   |
| <code>rebuild_PGN_Lasso</code>    | Logical. Whether to rebuild the peak-gene network via Lasso from scratch. If FALSE, the function will attempt to read from PGN_Lasso.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode.   |
| <code>rebuild_PGN_RF</code>       | Logical. Whether to rebuild the peak-gene network via random forest from scratch. If FALSE, the function will attempt to read from PGN_RF.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode.  |
| <code>rebuild_PGN_XGB</code>      | Logical. Whether to rebuild the peak-gene network via XGBoost from scratch. If FALSE, the function will attempt to read from PGN_XGB.mtx under dirpath/test in single mode or dirpath/state_name/test in compare mode.   |
| <code>relearn_pg_embedding</code> | Logical. Whether to relearn the low-dimensional representations for peaks and genes from scratch. If FALSE, the function will attempt to read from node_embeddings.mtx, node_used_peak.csv, node_used_gene.csv under dirpath/embedding in single mode or dirpath/state_name/embedding in compare mode. |

|                         |  |
|-------------------------|--|
| <code>save_file</code>  | Logical, whether to save the output to a file.   |
| <code>pg_method</code>  | The vector of methods used to construct peak-gene net. Default is c("Lasso", "RF", "XGBoost"). |
| <code>python_env</code> | Name or path of the Python environment to be used.   |

### Value

The scPOEM result.

**Single Mode** Returns a list containing the following elements:

- `E` Low-dimensional representations of peaks and genes.
- `peak_node` Peak IDs that are associated with other peaks or genes.
- `gene_node` Gene IDs that are associated with other peaks or genes.

**Compare Mode** Returns a list containing the following elements:

- `state1` name The single-mode result for the first condition.
- `state2` name The single-mode result for the second condition.
- `compare` A summary list containing:
  - `E_g2` Low-dimensional embedding representations of genes under the two conditions.
  - `common_genes` Genes shared between both conditions and used in the analysis.
  - `diffRegulation` A list of differential regulatory information for each gene.

### Examples

```
library(scPOEM)
library(monocle)
dirpath <- "./example_data"
# An example for analysing a single dataset.
# Download and read data.
data(example_data_single)
single_result <- scPOEM(mode = "single",
                         input_data=example_data_single,
                         dirpath=file.path(dirpath, "single"),
                         save_file=FALSE)

# An example for analysing and comparing datasets from two conditions.
# Download compare mode example data
data(example_data_compare)
compare_result <- scPOEM(mode = "compare",
                           input_data=example_data_compare,
                           dirpath=file.path(dirpath, "compare"),
                           save_file=FALSE)
```

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